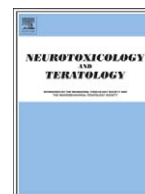




Contents lists available at ScienceDirect

Neurotoxicology and Teratology

journal homepage: www.elsevier.com/locate/neutera

Defining the neurotoxin derived illness chronic ciguatera using markers of chronic systemic inflammatory disturbances: A case/control study[☆]

Ritchie C. Shoemaker^{a,*}, Dennis House^a, James C. Ryan^b

^a Center for Research on Biotxin Associated Illnesses, Pocomoke, MD, USA

^b Marine Biotoxins Program, NOAA-National Ocean Service, Charleston, SC, USA

ARTICLE INFO

Article history:

Received 25 February 2010

Received in revised form 25 May 2010

Accepted 26 May 2010

Available online xxx

Keywords:

Ciguatera fish poisoning

Ciguatoxin

Inflammation

Immune

Biotoxin

Chronic inflammatory response syndrome

ABSTRACT

Background: Ciguatoxins are extremely potent neurotoxins, produced by tropical marine dinoflagellates, that persistently enter into our food web. Over 100,000 people annually experience acute ciguatera poisoning from consuming toxic fish. Roughly 5% of these victims will develop chronic ciguatera (CC), a widespread, multisymptom, multisystem, chronic illness that can last tens of years. CC is marked by disproportionate disability and non-specific refractory symptoms such as fatigue, cognitive deficits and pain, and is suggestive of other illnesses. Its unknown pathophysiology makes both diagnosis and treatment difficult.

Objectives: We wanted to compare objective parameters of visual contrast sensitivity testing, measures of innate immune response and genetic markers in cases to controls to assess the potential for the presence of persistent inflammatory parameters that are demonstrated in other biotoxin associated illnesses at a single specialty clinic.

Methods: Using 59 CC cases and 59 controls we present in retrospective review, in all cases, abnormalities in immune responses paralleling the chronic systemic inflammatory response syndrome seen in several other chronic diseases.

Results: This study defines a preliminary case definition using medical history, total symptoms, visual contrast sensitivity, HLA DR genotype analysis, reduction of regulatory neuropeptides VIP and MSH, and multiple measures of inflammatory immune response, especially C4 and TGFβ1, thereby providing a basis for identification and targeted therapy.

Conclusions: CC provides a model for chronic human illness associated with initiation of inflammatory responses by biologically produced neurotoxins.

© 2010 Published by Elsevier Inc.

Abbreviations: ACLA, anti-cardiolipin antibodies; ACTH, adrenocorticotrophic hormone; ADH, antidiuretic hormone; AGA, anti-gliadin antibodies; CBC, complete blood count; CC, chronic ciguatera; CIRS, chronic inflammatory response syndrome; CRP, C-reactive protein; CSM, cholestyramine; CTX, ciguatoxin; C3a, split activation product of C3; C4, fourth member of complement system; C4a, split activation product of C4; EAE, experimental autoimmune encephalitis; FACT, functional acuity contrast test (®); GGTP, gamma-glutamyl transpeptidase; HLA DR, Human leukocyte antigen Class II, DR locus; MASP, mannose binding lectin associated protease 2; MSH, alpha melanocyte stimulating hormone; MMP9, matrix metalloproteinase 9; PAI-1, plasminogen activation inhibitor-1; TGF, beta-1 transforming growth factor beta-1; T reg, T regulatory cell; TSH, thyroid stimulating hormone; VCS, visual contrast sensitivity; VEGF, vascular endothelial growth factor; VGSC, voltage gated sodium channel; VIP, vasoactive intestinal polypeptide; vWF, von Willebrand's profile.

[☆] NOAA disclaimer: This publication does not constitute an endorsement of any commercial product or intend to be an opinion beyond scientific or other results obtained by the National Oceanic and Atmospheric Administration (NOAA). No reference shall be made to NOAA, or this publication furnished by NOAA, to any advertising or sales promotion which would indicate or imply that NOAA recommends or endorses any proprietary product mentioned herein, or which has as its purpose an interest to cause the advertised product to be used or purchased because of this publication.

* Corresponding author. 500 Market Street Pocomoke City, MD 21851, USA. Fax: +1 410 957 3930.

E-mail address: ritchieshoemaker@msn.com (R.C. Shoemaker).

1. Introduction

Ciguatera, the most common marine poisoning worldwide, is acquired after ingestion of toxins produced by the tropical marine dinoflagellate *Gambierdiscus* spp. The most incriminated toxins in this illness are ciguatoxins (CTX), a suite of colorless, odorless, heat stable, cyclic polyether neurotoxins that are potent activators of voltage gated sodium channels. Ciguatoxins are biotransformed and biomagnified through trophic transfer in multiple fish species, and by recent estimates result in more than 100,000 cases annually of ciguatera fish poisoning, while considerable under-reporting still exists [17]. Ciguatoxin congeners from the Pacific and Indian Oceans, and the Caribbean Sea differ slightly in their structures and toxicities, which may underlie the variability of symptoms following exposure in these regions [27]. The predominant congener found in fish flesh of the Pacific, Pacific ciguatoxin 1 (P-CTX-1), can cause human illness at 0.1 ppb and is roughly 10 times more potent than the most common Caribbean congener, Caribbean ciguatoxin 1 (C-CTX-1) [26]. This extreme potency makes detection of ciguatoxins in fish difficult even in the most advanced research labs. Species of *Gambieridiscus* prefer to

live as epiphytes on macroalgae, which can now be found dominating newly bleached coral reefs. Given the acceleration of global coral bleaching, there are reasonable public health concerns regarding expansion of habitat for this toxic dinoflagellate. Elimination of ciguatoxins from fish is reported to be slow [26], which may serve as a reservoir for toxin accumulation in the food web.

Recognition of acute ciguatera poisoning is based on a history of near-immediate onset of a multisystem, multisymptom illness acquired after eating piscivorous reef fish, although many herbivorous fish are also toxic. Acute symptoms usually are (i) gastrointestinal, especially nausea, vomiting and diarrhea; (ii) neurologic, with numbness, tingling, paresthesias and dysesthesias; (iii) general symptoms of fatigue, weakness and peripheral pain [31]. Most patients recover without treatment. However, approximately 5% of patients develop a chronic illness, often lasting years, termed chronic ciguatera (CC) [37]. CC is characterized by persistent symptoms including fatigue, cognitive deficits, chronic pain and respiratory restriction, in addition to symptoms that may not resolve from the acute stage. The exact prevalence of CC cases is unknown due to the lack of a case definition, the inability to routinely associate the chronic illness with a distant point source exposure, and the potential for misdiagnosis as there are no diagnostic markers that would separate CC from other chronic disorders. The occurrence of ciguatera cases both in the tropics, where fresh fish may be consumed, and distant areas where ciguatoxic fish may be exported further confounds diagnosis. Without biomarkers or understanding the chronic syndrome's pathophysiology, chances for therapies targeted to specific mechanisms in CC are remote.

Currently, treatment of ciguatera is limited. Native islanders commonly turn to herbal remedies whose characterization is ongoing [7], but the efficacy of such therapy still remains to be determined. Although a double blinded trial showed no significant difference in outcome between saline and the osmolyte mannitol [44], use of mannitol in acute cases reportedly provides benefit if administered shortly after exposure. Likewise, little therapy exists for the chronic syndrome. Since 1999, the Center for Research on Biotxin Associated Illnesses (author affiliation, RS) has treated over 200 CC patients using cholestyramine (CSM), an orally administered, non-absorbable anion-binding resin as the first step in sequential therapies. Use of CSM consistently reduced symptoms when used as the initial therapy, although less benefit was seen in patients with illness of longer duration. Such persistent illness underscored the need for newer diagnostic modalities to achieve enhanced therapies.

As the mechanism of action for ciguatoxins is well characterized, the acute presentation is understood. However, the chronic syndrome appears more complex than the result of transient nervous injury and peripheral neuropathies. As literature on chronic inflammatory response syndromes emerged [1,8,16,25], many similarities to CC were noted. The current study employed newly available laboratory blood tests, especially the regulatory neuropeptides alpha melanocyte stimulating hormone (MSH) and vasoactive intestinal polypeptide (VIP); the split product of complement component C4, C4a; and transforming growth factor beta (TGF β 1) to determine if CC results from a complex dysregulation of innate and adaptive immune responses. We additionally sought to identify if HLA DR haplotypes, recently found predictive for other chronic illnesses of similar characteristics, were predictive of CC. A two tiered approach of (i) medical history, symptom rosters and visual contrast sensitivity (VCS) testing, coupled with (ii) lab testing of HLA DR, and multiple measures of immune response successfully identified cases of CC as markedly different from controls and other non-biotxin-associated, chronic illnesses such as asthma, somatoform disorder, allergy and depression.

Aspects of inflammatory pathways critical to the data were evaluated to advance understanding of how these pathways may contribute to the illness. This is the first paper to describe any underlying pathophysiology in the CC syndrome and is meant to

expose its elements for discussion, diagnosis, treatment and future work. The work is exploratory; it is a building block and not an absolute presentation of endpoints.

2. Methods

2.1. Patients

For all cases and controls, medical history was obtained concerning possible confounders, including, but not limited to other biotoxin exposure (other dinoflagellates, fungi, actinomycetes, mycobacteria, endotoxin-producing bacteria, cyanobacteria, apicomplexans and spirochetes), undiagnosed neurologic disease, alcoholism, occupational exposure to solvents, petroleum products, known neurotoxins and metal fumes. For cases, differential diagnosis techniques were used to determine whether or not a cause of illness other than ciguatera could be identified. Patients were included as CC cases (N=59) if they were considered to have (1) developed an acute illness typical of ciguatera following piscivorous reef fish consumption, (2) no confounding illnesses and (3) symptoms that persisted beyond three months. Confirmation of the presence of ciguatoxin by testing in fish was not required for diagnosis as such testing (i) is rarely readily available in all locations (ii) more often than not, there is no remaining fish after the meal or remaining fish has been discarded. Patients coming to the clinic for well-physicals were included as controls (N=59) if they had (i) no illness of any kind requiring acute intervention during that office visit; (ii) no history of acute multisystem illness after consumption of fish, or multisystem, multisymptom illness following exposure to environmentally produced biotoxins as described above; (3) any untreated chronic illness. Patients meeting inclusion criteria received a physical examination and blood analyses. Pregnant or nursing patients were excluded from study participation. All participants signed a HIPAA waiver permitting use of their clinical data. Internal review board (IRB) approval for retrospective analysis was obtained from the Copernicus Group IRB, Cary, NC. Participants were not remunerated for study participation.

2.2. Vision testing

Visual contrast sensitivity (VCS) testing measures the eye's ability to resolve patterns and was performed by an experienced physician using a previously published protocol [45]. Visual acuity and VCS testing were administered monocularly, with patients wearing any necessary corrective lenses, under a "daylight" illuminator (exceeding 70 fL) in a clinical unit with normal background lighting. A test card holder was used to position the acuity and VCS test cards at a constant, standardized distance (acuity – 36 cm, contrast sensitivity – 46 cm).

Visual acuity using Snellen score (e.g. 20/20) was determined for each eye using the acuity test card (MIS Pocket Vision Guide, © 1997 MIS, Inc.). To avoid confounding the VCS results, a visual acuity of 20:50 or better was required for each eye to be included in analysis. All participants had at least one eye included in analysis (N=112 in cases, N=113 in controls). Two-tailed Student t-tests were performed, using the mean score \pm s.e.m. of each participant's two eyes, to determine if acuity scores differed significantly (0.05) between cohorts.

The contrast sensitivity test card (Functional Acuity Contrast Test (FACT), Stereo Optical Co., Chicago, IL) contained a matrix (5 \times 9) of circles filled with sinusoidal gratings (dark and light bars) with spatial frequencies of 1.5, 3, 6, 12 and 18 cycles/ $^\circ$ of visual arc. The grating bars were oriented either vertically, or tilted 15 $^\circ$ to the left or right. Subjects identified the orientation of the grating by saying either: vertical, left, right or blank. The contrast sensitivity score for each row (spatial frequency) was recorded as the contrast of the last circle correctly identified on that row following verification by repeated testing of that circle. The procedure was repeated for each row in

193 descending order. The units of analysis for the VCS test were the mean
194 scores \pm s.e.m. of the participant's two eyes at each spatial frequency.

195 2.3. Blood tests

196 Laboratory measurements were performed by CLIA licensed
197 facilities, LabCorp, Quest Diagnostics, National Jewish Center and
198 Cambridge Biomedical. Testing included HLA DR by PCR, alpha
199 melanocyte stimulating hormone (MSH), vasoactive intestinal pep-
200 tide (VIP), leptin, matrix metalloproteinase 9 (MMP9), split product
201 of complement component 3 (C3a) and component 4 (C4a), trans-
202 forming growth factor beta-1 (TGF β 1), IgG for gliadin (AGA), and
203 IgM for cardiolipin (ACLA), vascular endothelial growth factor
204 (VEGF), plasminogen activator inhibitor (PAI-1), cortisol, erythrocyte
205 sedimentation rate, C-reactive protein (CRP), lipid profile, complete
206 blood count (CBC), comprehensive metabolic panel (CMP), gamma-
207 glutamyl transpeptidase (GGTP), thyroid stimulating hormone (TSH),
208 lipid profile, and von Willebrand's profile. Patients were classified
209 abnormal for von Willebrand's antigen for results either <50 or
210 >150 IU. Dysregulation of simultaneously measured ACTH/cortisol and
211 ADH/osmolality was determined by adding (i) the number of cases with
212 absolute high (ACTH >45 or cortisol >21 ; ADH >13 or osmolality >300)
213 or low (ACTH <5 or cortisol <4 ; ADH <1.3 or osmolality <275) values
214 for the two paired tests; to the cases (ii) in which ACTH was below 10
215 when cortisol was below 7; or ADH was below 2.2 when osmolality
216 was 292–300; to the cases (iii) in which ACTH was >15 when cortisol
217 was >16 ; and ADH >4.0 when osmolality was 275–278.

218 2.4. Statistical methods

219 There were 37 symptoms and 22 blood parameters measured in
220 this study for a total of 59 variables not including VCS. Because of
221 this multiplicity problem, the Bonferroni correction was applied
222 to symptom and blood variables which resulted in a single variable
223 p-value being considered statistically significant if $p<001$ (.05/59
224 rounded) in order to have an experiment wise $p<05$. The units of
225 analysis for the VCS test were the mean scores of the participant's
226 two eyes at each spatial frequency. The VCS data were analyzed
227 using multivariate analyses of variance (MANOVA, with the Wilks'
228 lambda statistic) procedures suitable for repeated measures with an
229 $\alpha=0.05$. The factors in this model were group, spatial frequency,
230 age and their interaction terms. A factor for gender was not included,
231 as no gender differences in susceptibility to ciguatoxin-induced
232 effects had been indicated, and no gender differences in VCS have
233 been reported. Results further showed that a significant group-by-
234 spatial-frequency interaction were further analyzed in step down,
235 two-tailed Student's t-tests ($\alpha=0.05$), the equivalent of a univariate
236 ANOVA, to determine which spatial frequencies accounted for the
237 overall effect.

238 2.4.1. Symptoms

239 The prevalence of each symptom in the illness and control groups
240 was compared for statistical significance ($p<0.001$) using Fisher's
241 exact test.

242 2.4.2. Blood testing parameters

243 For each blood parameter, the difference between the two groups
244 was tested for statistical significance ($p<0.001$) using the two-tailed
245 two-sample Student t-test.

246 2.4.3. Statistical program

247 JMB of SAS was used for data analysis.

248 2.4.4. VCS

249 The VCS data were analyzed using multivariate analyses of
250 variance procedures suitable for repeated measures. The factors in

the model were group, spatial frequency, and their interaction. A
251 significant ($p<0.05$) overall group-by-spatial-frequency interaction
252 was further analyzed by a two-tailed Student t-test at each spatial
253 frequency to determine which frequencies accounted for the effect. 254

254 2.4.5. HLA haplotype relative risk

255 Differences in relative risk were assessed using incidence in cases
256 to incidence in an established control population (N=111) [46].
257 Results were considered significant if the ratio exceeded 2.0. 258

259 3. Results

260 3.1. Patient demographics

261 The 118 patients were predominately Caucasian with five African
262 Americans (2 cases, 3 controls) and two Asian Americans (1 case, 1
263 control). Putative cases were selected from patients seeking therapy
264 for a chronic illness and meeting inclusion criteria described in
265 *Methods*. Based on location for acquisition of illness (*Supp. Table 1*)
266 we feel the majority of cases were exposed to the less potent
267 Caribbean ciguatoxin. Mean age was 51 years for the 39 female
268 and 20 male controls, and 50 years for the 17 female and 42 male
269 cases. Although gender was not evenly matched, diagnostic facilities
270 (LabCorp, etc.) directly communicated there are no known gender
271 differences in normative values for blood tests presented here.
272 Additionally, comparisons were broken out by gender (*Supp. Table 2*)
273 with similar results.

274 3.2. Symptom roster

275 Patients identified the presence or absence of 37 different
276 symptoms (*Table 1*). For all symptoms except sinus congestion and
277 joint pain the prevalence among cases was significantly higher than
278 controls ($p<001$). The greatest separation (occurrence of symptom in
279 cases minus occurrence in controls) was seen for light sensitivity,
280 memory impairment, and fatigue; while the greatest sensitivity
281 (occurrence in cases divided by occurrence in controls) was seen for
282 unusual pain, cramping, ice pick pain, and confusion. Controls had few
283 of the queried symptoms, but paralleled those seen in control groups
284 for similar studies [47,48].

285 3.3. Visual testing

286 Visual contrast deficits have been shown after a number of neu-
287 rologic insults such as mercury exposure [12], Parkinson's disease [9],
288 and organic solvent exposure [19] among others. No significant
289 differences were noted between cases and controls for visual acuity.
290 However, cases showed a significant ($p<0.005$) pattern of depressed
291 VCS at all frequencies with a maximal shift from 6 cycles/ $^{\circ}$ of visual arc
292 to 3 cycles/ $^{\circ}$ of visual arc (*Fig. 1*).

293 3.4. Blood tests

294 Not all patients were subjected to the complete battery of diag-
295 nostic blood tests as severity of immune dysfunction was only
296 apparent after analysis of the first 30 cases. Increased relative risk
297 (>2.0) was seen for three haplotypes of HLA DR: 1) DRB1–4, DQ-3 and
298 DRB4–53; 2) DRB1–4, DQ-7/8 and DRB4–53; and 3) DRB1–11, DQ-3,
299 DRB3–52B (*Supp. Table 3*). Statistical differences ($p<0.001$) were
300 seen between cases and controls for (i) serum protein measures of
301 the immune parameters; alpha melanocyte stimulating hormone
302 (MSH), vasoactive intestinal peptide (VIP), matrix metalloproteinase
303 9 (MMP9), split product of complement component 4 (C4a), trans-
304 forming growth factor beta-1 (TGF β 1); (ii) autoimmune param-
305 eters of IgG for gliadin (AGA), and IgM for cardiolipin (ACLA);
306 (iii) clotting parameters of von Willebrand's profile and (iv) hormone

Table 1
Symptom roster. The number of cases and controls reporting symptoms by group.

Symptom	Group			p-value ^a
	Control (n = 59)	Ciguatera (n = 59)		
Fatigue	11	56		<.001
Weak	5	44		<.001
Ache	5	45		<.001
Cramp	1	39		<.001
Unusual pain	0	35		<.001
Ice pick pain	1	37		<.001
Headache	11	47		<.001
Light sensitivity	5	51		<.001
Red eyes	3	29		<.001
Blurred vision	5	37		<.001
Tearing	6	29		<.001
Sinus	19	32		0.008
Cough	11	30		<.001
Shortness of breath	8	45		<.001
Abdominal pain	8	41		<.001
Diarrhea	3	42		<.001
Joint pain	16	31		0.003
Morning stiffness	2	24		<.001
Memory	7	52		<.001
Focus/concentration	3	45		<.001
Word recall	5	40		<.001
Decrease assimilation	4	35		<.001
Confusion	1	37		<.001
Disorientation	1	22		<.001
Skin sensitivity	1	27		<.001
Mood swings	4	43		<.001
Appetite	2	29		<.001
Sweats	3	33		<.001
Temp regulation	6	33		<.001
Thirst	5	33		<.001
Increased urination	4	34		<.001
Static shocks	1	28		<.001
Numbness	4	33		<.001
Tingling	5	43		<.001
Vertigo	4	29		<.001
Metallic taste	1	31		<.001
Tremor	1	15		<.001

^a From testing hypothesis of no difference between groups.

relationships of ACTH compared to simultaneously measured cortisol (ACTH/cortisol), and comparison of ADH to simultaneously measured osmolality (ADH/osmolality) as described in **Methods** (Table 2). No significant differences between cases and controls were seen for C3a,

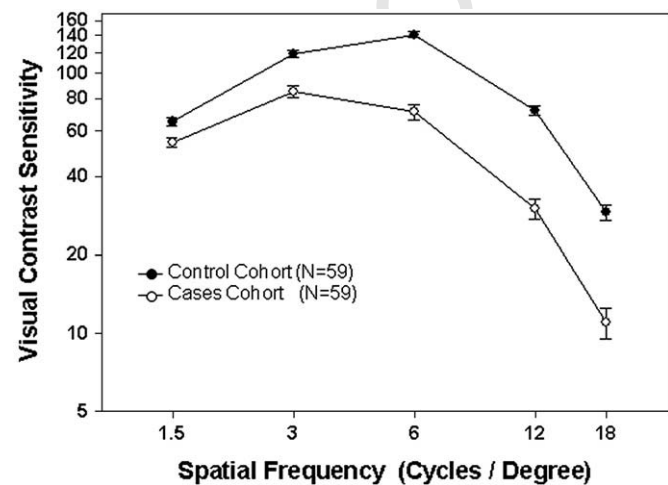


Fig. 1. Mean visual contrast sensitivity by spatial frequency show a comparison of visual contrast sensitivity between age matched controls and chronic ciguatera cases. The values of the spatial frequencies score are means of right and left eyes. Error bars indicate standard error. Significant ($p < 0.005$) differences were noted at all frequencies.

VEGF, PAI-1, ACTH, cortisol, erythrocyte sedimentation rate, CRP, lipid profile, CBC, CMP, GGTP, TSH or leptin. Of note is that cases presented with a bimodal distribution of VEGF with deficiency (<31) or elevation (>86) seen in 31 of 43 patients compared to 10 of 49 controls. These laboratory abnormalities identify a complex syndrome marked by host responses of inflammation, autoimmunity and coagulopathy.

Use of a two tiered structure for case definition served to separate all CC cases from all controls. All cases and no controls had the presence of a multisymptom illness from at least four organ systems, without confounders, persisting following consumption of a fish meal. Analysis of testing showed all 59 cases but no control subject presented with at least 4 of nine objective parameters including VCS deficits, HLA DR from a roster with a relative risk that exceeded 2.0, MSH, VIP, C4a, TGFβ1, MMP9, ACTH/cortisol and ADH/osmolality abnormalities (Table 3a). Not all patients were subjected to the complete battery of diagnostic blood tests as severity of immune dysfunction was only apparent after analysis of the first 30 cases. Although we could only include roughly half the study subjects for markers of TGFβ1, C4a and VIP, adding these tests aided case definition in mean number and distribution of abnormalities (Table 3b). The values given for case definition threshold in Table 3a are upper limit normative ranges defined by the testing facility (Quest, LabCorp, etc.).

4. Discussion

The results of this study identify, for the first time, that a series of immune abnormalities are present in chronic ciguatera cases. Objective identification of these abnormalities can now be routinely performed and should speed the advent of treatments for victims who may suffer with this syndrome for years. The discussion of these data and abnormalities involves various biochemical pathways and mechanisms, not always in a confluent format, but valuable to understanding the pathophysiological backdrop.

Table 2
Individual study parameters. The mean and standard deviation results of cases and controls by group, with N subjects for each test.

Variable	Controls			Cases			p-value ^a
	Mean	N1	s.d.	Mean	N2	s.d.	
Age	51.1	59	12.1	50.7	59	14.4	0.890
Total number Of Symptoms	3.1	59	2.3	22.4	59	7.4	<.0001
VIP	35.5	8	10.7	7.1	24	5.9	<.0001
MSH	34.9	57	12.2	9.8	59	6.7	<.0001
Leptin	18.3	55	26.3	13.5	53	15.8	0.250
ADH	3.9	52	2.0	1.6	54	1.5	<.0001
Osmo	290.2	52	4.8	296.8	53	10.1	<.0001
ACTH	22.6	53	12.5	26.1	47	30.1	0.436
Cortisol	15.5	54	18.3	16.6	46	9.1	0.715
CRP	2.2	51	2.8	2.1	52	2.4	0.728
MMP-9	266	59	138	510	51	314	<.0001
PAI-1	5.5	53	6.0	10.3	43	14.6	0.033
VEGF	70.2	46	38.8	69.0	42	98.1	0.943
IgE	45.9	51	67.9	62.7	27	87.5	0.350
TSH	2.5	53	1.5	2.1	32	1.5	0.257
vWF	0.08	13	0.28	0.64	23	0.58	<.0001
ACLA-IgA	0.02	58	0.13	0.06	34	0.24	0.883
ACLA-IgM	0.05	57	0.23	0.28	40	0.45	0.002
ACLA-IgG	0.04	57	0.19	0.03	34	0.17	0.885
AGA-IgA	0.04	57	0.19	0.11	35	0.32	0.138
AGA-IgG	0.05	57	0.23	0.47	36	0.51	<.0001
C3a	258	39	181	328	29	250	0.184
C4a	2324	41	1212	10,640	29	5859	<.0001
TGFβ-1	2076	12	1011	8296	19	4535	<.0001

^a From testing hypothesis of no difference between groups.

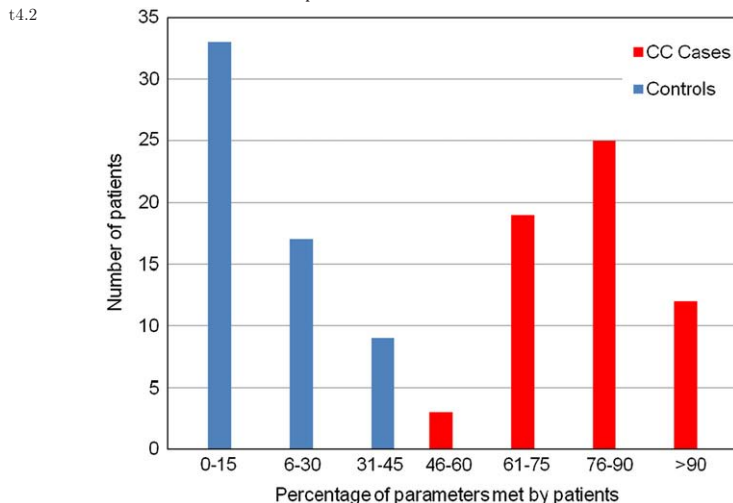
t3.1 **Table 3a**
Case definition parameters.

t3.2	Parameters	% + Cases (n = 59)	% + Controls (n = 59)
t3.3	VCS deficit	96	2
t3.4	HLA DR RR > 2	61	26
t3.5	MSH < 25 pg/mL	95	13
t3.6	VIP < 23 pg/mL	96	0
t3.7	C4a > 2830 ng/mL	89	25
t3.8	TGFβ-1 > 2380 pg/mL	89	25
t3.9	MMP9 > 332 ng/mL	61	22
t3.10	ADH/osmo dysregulation	83	14
t3.11	ACTH/cortisol dysregulation	54	13

t4.1 4.1. VGSC

t4.2 Ciguatoxins are extremely potent voltage gated sodium channel (VGSC) activators, exerting their acute effects predominantly on the peripheral nervous system. Although this mechanism of action explains the acute neuropathies acquired after exposure, the physiologic basis for the chronic syndrome was largely unapparent. VGSCs have now been characterized in several types of non-excitabile cells and studies have shown that these channels contribute to activation of inflammatory pathways in many immune cells [42]. In a study of microglial and macrophage activation, Craner et al. [13] demonstrated correlation between up-regulation of VGSC 1.6 and transition from resting state to activated phenotypes in MS and experimental autoimmune encephalitis (EAE). Moreover, they demonstrated the utility of phenytoin, a VGSC blocker, in mitigation of EAE. Carrithers et al. demonstrated VGSC 1.5 plays an active role in macrophage endosomal acidification and phagocytosis, an important component of antigen processing by dendritic cells. The authors posit that hyperacidification through VGSC 1.5 activation in macrophages is comparable to that seen in cystic fibrosis, and further suggest, this mechanism may play a role in chronic infections and autoimmune disease [10]. VGSCs were also found to regulate invasive/motile properties in Jurkat cells, a T-cell line [18]. Brevetoxin, another dinoflagellate toxin, similar to ciguatoxin in structure and identical in mode of action, was found localized to macrophages and lymphocytes in natural exposures of manatees [6]. During preparation of this manuscript, a new study revealed that exposure of macrophages to P-CTX-1 elicited a response similar to that of lipopolysaccharides at the mRNA level [30]. Exposure to ciguatoxins may have a bimodal effect, quickly damaging sensitive neurons, while also generating

t4.1 **Table 3b**
Distribution of case definition parameters.



highly activated immune cells. Damaged cells can release a class of endogenous pro-inflammatory molecules termed alarmins, which then initiate both innate and adaptive immune responses to aid in repair and removal of injured tissue [35]. Alarmins have been shown to interact with Toll-like receptors, classical receptors for initiating an innate immune response [4]. A genomic study in liver of acute ciguatoxin exposure in mice showed several alarmin (defensins/cryptidins) genes were up-regulated at 4 and 24 h post toxin exposure [33]. In whole blood of these animals, significant immune system activation was seen, the authors citing the data set had many genes known to be important in allergic asthma models, although the gene expression was confounded by the rodent's hypothermic response to toxin [43].

4.2. C3a, C4a

The complement system is a component of both innate and adaptive immune responses. Patients in this cohort had four times the upper limit normative value for the anaphylatoxin C4a, although near normal levels of C3a, a product just downstream of C4 activation. Complement activation through both the classical pathway and mannose binding lectin system will generate increased levels of C4a. Autoactivation of the C4 protease MASP2 has been reported, which could lead to persistent elevation of C4a [54]. The diversity of C4 genotypes can also influence disease progression [39]. The C4 gene locus resides in the HLA class III region and aside from simple polymorphic bases, this gene typically varies in diploid copy number (2-6, although >6 in rare instances), size (long and short forms), and isotypes (A and B, for acidic and basic) [5]. In the Caucasian population the maximal gene dosage of 6 has a frequency of 3.3% [5], a proportion not dissimilar to progression of acute to chronic ciguatera. Recent studies have shown interaction between the complement and coagulation systems with evidence of shared inhibitors and activators [1]. In particular, platelets and platelet microparticles can activate C4 in the absence of immune complexes [38].

4.3. Von Willebrand's profile

Results of von Willebrand's profile for CC patients show that the acute phase reactant Factor VIII remains abnormal, as does ristocetin associated cofactor and von Willebrand's antigen itself. Although unexplained bleeding is rare, disturbances in coagulation pathways are commonly seen in CC patients, just as in other chronic inflammatory response syndromes [41]. These protein level abnormalities would most likely result in abnormal coagulation times, although this theory has never been clinically tested for CC.

4.4. Neuropeptides VIP and MSH

The regularly observed deficits in two neuropeptides, vasoactive intestinal peptide (VIP) and melanocyte stimulation hormone (MSH), both neuroendocrine regulators of inflammatory responses, suggests an absence of regulation of inflammation in the development and persistence of CC. These two neuropeptides have profound anti-inflammatory effects both *in vivo* and *in vitro*; each shows great promise for treatment of inflammatory disease progression (for excellent reviews see [8,16]). Deficiency in these neuropeptides can be acquired either acutely or delayed, as well as through diverse mechanisms such as acute brain injuries [28] or persistent viral infection [51]. Although both VIP and MSH have specific receptors in immune cells, MSH is also thought to directly antagonize the classic inflammatory interleukin-1β receptor [34]. Receptor density and affinity for these peptides have been proven crucial to function. VIP can enhance newly defined inflammatory Th17 differentiation pathways through VIP receptor type 1 (VPAC1) [55] while VPAC2 levels are critical in maintaining Th1 and Th2 states in CD4+ T cells of MS

patients [50]. Furthermore, VIP receptor agonists in rats showed efficacy in protection against Alzheimers related learning impairment [21] while deficiency was shown to cause cognitive deficits in mice [11], a common symptom of cases in this cohort. Another critical role for these neuropeptides is the induction of tolerogenic dendritic cells and generation of T regulatory cells (Tregs), which suppress autoreactive T cells and autoimmune progression [16]. Even in healthy individuals autoreactive T cells can escape clonal deletion and must be policed in the periphery by Tregs to prevent pathologic autoimmunity [14]. Of note, these CC study patients with deficiency of VIP and MSH show evidence of autoimmune findings in elevated anti-gliadin and anti-cardiolipin antibodies.

4.5. TGFβ1

This cytokine has wide ranging effects on the immune system, including important roles in autoimmune and inflammatory disease. However, timing, duration and target tissue are important aspects for its protection or pathological activity, so the effects of TGFβ1 elevation seen in CC have yet to be fully understood. Similar to VIP and MSH, TGFβ1 can regulate T-cell differentiation pathways and is considered anti-inflammatory [25]. Matrix metalloproteinase-9 (MMP9), whose expression is up-regulated by TGFβ1, can influence disease progression by both tissue destruction and cytokine processing and its elevation is also characteristic of many inflammatory and autoimmune conditions [53]. Elevated levels of both MMP9 and TGFβ1 have been reported in systemic sclerosis, a generalized disorder of the microvasculature characterized by excessive fibrosis [40]. Further, the role of TGFβ1 as a stimulant to pro-fibrotic effects in lung parenchyma, including epithelial to mesenchymal transformation, may support an explanation of restrictive pulmonary function seen in these current cases [29] (data not shown).

The incidence of developing CC from acute ciguatera remains relatively low (5%) and it is unknown as to what generates this transition. Curiously, the incidence of chronic ciguatera parallels the incidence of particular HLA haplotypes (Supp. Table 3). Although a relatively small cohort size for HLA allele analysis, cases showed an increased relative risk for certain immune haplotypes. Findings of increased risk associated with HLA DRB1–4 in chronic disease, as seen in this cohort, are not uncommon and were also seen in patients with persistent illness from Lyme disease [49], water damaged building cases [46], autoimmune hepatitis [32], severe malaria [36], pulmonary tuberculosis [22] and rheumatoid arthritis [20], among other illnesses. Additionally, expression of HLA DR genes in antigen presenting cells can be regulated by cytokine and Th1/Th2 ratio [24], parameters that are influenced by the immunomodulators already discussed.

Studies of chronic ciguatera (CC) are largely anecdotal case reports, with recording of symptoms but without laboratory testing. Given that many chronic medical conditions may present with similar symptoms, especially those for which objective diagnostic laboratory parameters have not been defined, greater accuracy in diagnosis of CC would be provided by a case definition that includes objective lab testing. Use of symptom recording in a medical history followed by visual contrast sensitivity (VCS) testing is inexpensive and rapid, making these measures ideally suited for screening large numbers of patients, especially in endemic areas of the Caribbean and South Pacific. When combined, an episode of reef fish consumption, presence of symptoms from four organ systems and VCS deficits, with an absence of other known biotoxin exposures, identified 57/59 CC patients. Given that the objective lab parameters identified all 59 CC cases, but results may take 30 days, the use of symptom clusters and VCS could provide a non-invasive, rapid and reliable on-site screening tool that allows immediate therapy while specific blood labs are run. The later-arriving lab results will guide subsequent therapies beyond initial intervention using CSM, understanding that resolution of CC requires reduction of symptoms, correction of VCS deficits and

correction of elevated MMP9, C4 and TGF beta-1. What remains unexplored in CC is the effect of repeated sub-acute exposures, as repeat exposures in other biotoxin associated illnesses lead to enhanced C4a and TGF bet-1 response and not reduction of response (Shoemaker 2005, 2006). As long as 25 years ago, measurable ciguatoxin was reported in most reef fish of endemic areas of French Polynesia [3] while more recently, a study of barracuda from the Florida Keys showed 60% were positive for ciguatoxins [15]. Interestingly, a ciguatoxin like epitope was identified in CFS patients using a monoclonal antibody [23].

5. Conclusions

These data support a complex interaction of environmental exposure, genetics, innate and adaptive immunity, and neuropeptide regulatory mechanisms in patients with CC. Along with abnormal neuropeptide regulation, increased autoimmune findings and genetic susceptibility we document increased levels of the inflammatory mediators TGF β1, MMP9 and C4a. These findings are consistent with the hypothesis that CC is an illness characterized by immune dysregulation based on genetic control of host responses. Further research is required. This immune dysregulation seen in CC parallels that seen in other chronic inflammatory response syndromes (CIRS) initiated in diverse diseases such as sepsis [41], acute liver failure [2] and acute multiple trauma [52]. The differences in self-limited acute illness versus development of chronic illness may be related to HLA genotype, dysregulation of antigen presentation or policing of autoreactive T cells as seen in some of the above diseases. Now that such abnormalities are known to be routinely found in CC cases, these markers not only help identify the illness but also provide a basis for targeted therapies and monitoring of sequential intervention. As research of the chronic illness caused by ciguatoxins expands, additional delineation of the physiological basis of fatigue, cognitive, neurologic, rheumatologic, respiratory, and other symptoms may permit sub-typing of cases, leading to improved therapies.

Conflict of interest statement

RS has provided testimony in litigation regarding ciguatera. Ritchie C. Shoemaker MD is a medical director of the Center for Research on Biotoxin Associated Illnesses, which is a not-for-profit institution in the United States. The Center for Research on Biotoxin Associated Illnesses funded the work carried out in this research paper.

Acknowledgements

The authors would like to thank F. Van Dolah, Y. Bottein, T. Leighfield and K. Burnett for their review of this manuscript. Grant information: Funding for this study was provided by the Center for Research on Biotoxin Associated Illnesses, a private, non-profit 501-c-3 organization, Pocomoke, MD 21851.

Appendix A. Supplementary data

Supplementary data associated with this article can be found, in the online version, at doi:10.1016/j.ntt.2010.05.007.

References

- U. Amara, D. Rittirsch, M. Flierl, U. Bruckner, A. Klos, F. Gebhard, J.D. Lambris, M. Huber-Lang, Interaction between the coagulation and complement system, *Adv. Exp. Med. Biol.* 632 (2008) 71–79.
- C.G. Antoniadis, P.A. Berry, J.A. Wendon, D. Vergani, The importance of immune dysfunction in determining outcome in acute liver failure, *J. Hepatol.* 49 (2008) 845–861.
- R. Bagnis, Ciguatera, sante et developpement en Polynesie francaise, *Unión Méd. Can.* 115 (1986) 502–506.

- 555 [4] M.E. Bianchi, DAMPs, PAMPs and alarmins: all we need to know about danger, *J.*
556 *Leukoc. Biol.* 81 (2007) 1–5.
- 557 [5] C.A. Blanchong, E.K. Chung, K.L. Rupert, Y. Yang, Z. Yang, B. Zhou, J.M. Moulds, C.Y.
558 Yu, Genetic, structural and functional diversities of human complement
559 components C4A and C4B and their mouse homologues, Slp and C4, *Int.*
560 *Immunopharmacol.* 1 (2001) 365–392.
- 561 [6] G.D. Bossart, D.G. Baden, R.Y. Ewing, B. Roberts, S.D. Wright, Brevetoxicosis in
562 manatees (*Trichechus manatus latirostris*) from the 1996 epizootic: gross,
563 histologic, and immunohistochemical features, *Toxicol. Pathol.* 26 (1998)
564 276–282.
- 565 [7] R. Boydrone-Le Garrec, E. Benoit, M.P. Sauviat, R.J. Lewis, J. Molgo, D. Laurent,
566 Ability of some plant extracts, traditionally used to treat ciguatera fish poisoning,
567 to prevent the in vitro neurotoxicity produced by sodium channel activators,
568 *Toxicol.* 46 (2005) 625–634.
- 569 [8] T. Brzoska, T.A. Luger, C. Maaser, C. Abels, M. Bohm, Alpha-melanocyte-
570 stimulating hormone and related tripeptides: biochemistry, antiinflammatory
571 and protective effects in vitro and in vivo, and future perspectives for the
572 treatment of immune-mediated inflammatory diseases, *Endocr. Rev.* 29 (2008)
573 581–602.
- 574 [9] C. Bulens, J.D. Meerwaldt, G.J. Van der Wildt, Effect of stimulus orientation on
575 contrast sensitivity in Parkinson's disease, *Neurology* 38 (1988) 76–81.
- 576 [10] M.D. Carrithers, S. Dib-Hajj, L.M. Carrithers, G. Tokmoulina, M. Pypaert, E.A. Jonas,
577 S.G. Waxman, Expression of the voltage-gated sodium channel Nav1.5 in the
578 macrophage late endosome regulates endosomal acidification, *J. Immunol.* 178
579 (2007) 7822–7832.
- 580 [11] D. Chaudhury, D.H. Loh, J.M. Dragich, A. Hagopian, C.S. Colwell, Select cognitive
581 deficits in vasoactive intestinal peptide deficient mice, *BMC Neurosci.* 9 (2008) 63.
- 582 [12] M.F. Costa, S. Tomaz, J.M. de Souza, L.C. Silveira, D.F. Ventura, Electrophysiological
583 evidence for impairment of contrast sensitivity in mercury vapor occupational
584 intoxication, *Environ. Res.* 107 (2008) 132–138.
- 585 [13] M.J. Craner, T.G. Damarjian, S. Liu, B.C. Hains, A.C. Lo, J.A. Black, J. Newcombe, M.L.
586 Cuzner, S.G. Waxman, Sodium channels contribute to microglia/macrophage
587 activation and function in EAE and MS, *Glia* 49 (2005) 220–229.
- 588 [14] N.A. Danke, D.M. Koelle, C. Yee, S. Beheray, W.W. Kwok, Autoreactive T cells
589 in healthy individuals, *J. Immunol.* 172 (2004) 5967–5972.
- 590 [15] M.Y. Dechraoui, J.A. Tiedeken, R. Persad, Z. Wang, H.R. Granade, R.W. Dickey, J.S.
591 Ramsdell, Use of two detection methods to discriminate ciguatoxins from
592 brevetoxins: application to great barracuda from Florida Keys, *Toxicol.* 46
593 (2005) 261–270.
- 594 [16] M. Delgado, D. Ganea, Anti-inflammatory neuropeptides: a new class of
595 endogenous immunoregulatory agents, *Brain Behav. Immun.* 22 (2008)
596 1146–1151.
- 597 [17] L.E. Fleming, K. Broad, A. Clement, E. Dewailly, S. Elmir, A. Knap, S.A. Pomponi, S.
598 Smith, H. Solo Gabriele, P. Walsh, Oceans and human health: emerging public
599 health risks in the marine environment, *Mar. Pollut. Bull.* 53 (2006) 545–560.
- 600 [18] S.P. Fraser, J.K.J. Diss, L.J. Lloyd, F. Pani, A.-M. Chionio, A.J.T. George, M.B.A. Djamgoz,
601 T-lymphocyte invasiveness: control by voltage-gated Na⁺ channel activity, *FEBS*
602 *Lett.* 569 (2004) 191–194.
- 603 [19] Y. Gong, R. Kishi, S. Kasai, Y. Katakura, K. Fujiwara, T. Umemura, T. Kondo, T. Sato,
604 F. Sata, E. Tsukishima, et al., Visual dysfunction in workers exposed to a mixture
605 of organic solvents, *Neurotoxicology* 24 (2003) 703–710.
- 606 [20] M.A. Gonzalez-Gay, C. Garcia-Porrúa, A.H. Hajeer, Influence of human leukocyte
607 antigen-DRB1 on the susceptibility and severity of rheumatoid arthritis, *Semin.*
608 *Arthritis Rheum.* 31 (2002) 355–360.
- 609 [21] I. Gozes, A. Bardea, M. Bechar, O. Pearl, A. Reshef, R. Zamostiano, A. Davidson, S.
610 Rubinraut, E. Giladi, M. Fridkin, et al., Neuropeptides and neuronal survival:
611 neuroprotective strategy for Alzheimer's disease, *Ann. NY Acad. Sci.* 814 (1997)
612 161–166.
- 613 [22] E.I. Harfouch-Hammoud, N.A. Daher, Susceptibility to and severity of tuberculosis
614 is genetically controlled by human leukocyte antigens, *Saudi Med. J.* 29 (2008)
615 1625–1629.
- 616 [23] Y. Hokama, G.A. Uto, N.A. Palafox, D. Enlander, E. Jordan, A. Cocchetto, Chronic
617 phase lipids in sera of chronic fatigue syndrome (CFS), chronic ciguatera fish
618 poisoning (CCFP), hepatitis B, and cancer with antigenic epitope resembling
619 ciguatoxin, as assessed with MAb-CTX, *J. Clin. Lab. Anal.* 17 (2003) 132–139.
- 620 [24] Y. Imamura, T. Yokoyama, Y. Takesue, E. Hiyama, T. Sueda, The TH1/TH2 ratio in
621 patients with diminished expression of HLA-DR on monocytes, *Int. Congress Ser.*
622 1255 (2003) 225–229.
- 623 [25] M.A. Kriegl, M.O. Li, S. Sanjabi, Y.Y. Wan, R.A. Flavell, Transforming growth factor-
624 beta: recent advances on its role in immune tolerance, *Curr. Rheumatol. Rep.*
625 8 (2006) 138–144.
- 626 [26] L. Lehane, R.J. Lewis, Ciguatera: recent advances but the risk remains, *Int. J. Food*
627 *Microbiol.* 61 (2000) 91–125.
- 628 [27] R.J. Lewis, The changing face of ciguatera, *Toxicol.* 39 (2001) 97–106.
- 629 [28] S. Magnoni, N. Stocchetti, G. Colombo, A. Carlini, A. Colombo, J.M. Lipton, A. Catania,
630 Alpha-melanocyte-stimulating hormone is decreased in plasma of patients with
631 acute brain injury, *J. Neurotrauma* 20 (2003) 251–260.
- 632 [29] T. Makinde, R.F. Murphy, D.K. Agrawal, The regulatory role of TGF- β in airway
633 remodeling in asthma, *Immunol. Cell Biol.* 85 (2007) 348–356.
- 634 [30] M. Matsui, S. Kumar-Roine, H.T. Darius, M. Chinain, D. Laurent, S. Pauillac, Pacific
635 ciguatoxin 1B-induced modulation of inflammatory mediators in a murine
636 macrophage cell line, *Toxicol. In Press*, Corrected Proof.
- 637 [31] J. Matta, J. Navas, M. Milad, R. Manger, A. Hupka, T. Frazer, A pilot study for the
638 detection of acute ciguatera intoxication in human blood, *J. Toxicol. Clin. Toxicol.*
639 40 (2002) 49–57.
- 640 [32] A.J. Montano-Loza, H.A. Carpenter, A.J. Czaja, Clinical significance of HLA DRB103-
641 DRB104 in type 1 autoimmune hepatitis, *Liver Int.* 26 (2006) 1201–1208.
- 642 [33] J.S. Morey, J.C. Ryan, M.Y. Bottein Dechraoui, A.H. Rezvani, E.D. Levin, C.J. Gordon,
643 J.S. Ramsdell, F.M. Van Dolah, Liver genomic responses to ciguatoxin: evidence
644 for activation of phase I and phase II detoxification pathways following an acute
645 hypothermic response in mice, *Toxicol. Sci.* 103 (2008) 298–310.
- 646 [34] K.G. Mugridge, M. Perretti, P. Ghiara, L. Parente, Alpha-melanocyte-stimulating
647 hormone reduces interleukin-1 beta effects on rat stomach preparations possibly
648 through interference with a type I receptor, *Eur. J. Pharmacol.* 197 (1991)
649 151–155.
- 650 [35] J. Oppenheim, P. Tewary, G. de la Rosa, D. Yang, Alarmins initiate host defense, in:
651 M. Shurin, Y. Smolkin (Eds.), *Immune-mediated Diseases*, Springer New York,
652 New York, 2007, pp. 185–194.
- 653 [36] A.D. Osafo-Addo, K.A. Koram, A.R. Oduro, M. Wilson, A. Hodgson, W.O. Rogers,
654 HLA-DRB1*04 allele is associated with severe malaria in northern Ghana, *Am. J.*
655 *Trop. Med. Hyg.* 78 (2008) 251–255.
- 656 [37] J. Pearn, Models for the chronic fatigue syndrome: channelopathies and ciguatera,
657 in: T.K. Robers (Ed.), *The Clinical and Scientific Basis of Chronic Fatigue Syndrome:*
658 *From Myth towards Management*, 1998, Sydney.
- 659 [38] E.I. Peerschke, W. Yin, B. Ghebrehwiet, Platelet mediated complement activation,
660 *Adv. Exp. Med. Biol.* 632 (2008) 81–91.
- 661 [39] R. Porter, Complement polymorphism, the major histocompatibility complex and
662 associated diseases: a speculation, *Mol. Biol. Med.* 1 (1983) 161–168.
- 663 [40] M. Ram, Y. Sherer, Y. Shoenfeld, Matrix metalloproteinase-9 and autoimmune
664 diseases, *J. Clin. Immunol.* 26 (2006) 299–307.
- 665 [41] D. Rittirsch, M.A. Flierl, P.A. Ward, Harmful molecular mechanisms in sepsis, *Nat.*
666 *Rev. Immunol.* 8 (2008) 776–787.
- 667 [42] F. Roselli, P. Livrea, E. Jirillo, Voltage-gated sodium channel blockers as
668 immunomodulators, *Recent Pat. CNS Drug Discov.* 1 (2006) 83–91.
- 669 [43] J.C. Ryan, M.Y. Bottein Dechraoui, J.S. Morey, A. Rezvani, E.D. Levin, C.J. Gordon, J.S.
670 Ramsdell, F.M. Van Dolah, Transcriptional profiling of whole blood and serum
671 protein analysis of mice exposed to the neurotoxin Pacific Ciguatoxin-1,
672 *Neurotoxicology* 28 (2007) 1099–1109.
- 673 [44] H. Schnorf, M. Taurarii, T. Cundy, Ciguatera fish poisoning: a double-blind
674 randomized trial of mannitol therapy, *Neurology* 58 (2002) 873–880.
- 675 [45] R.C. Shoemaker, Residential and recreational acquisition of possible estuary-
676 associated syndrome: a new approach to successful diagnosis and treatment,
677 *Environ. Health Perspect.* 109 (Suppl 5) (2001) 791–796.
- 678 [46] R.C. Shoemaker, in: E. Johanning (Ed.), *Sick Building Syndrome in Water-damaged*
679 *Buildings: Generalization of the Chronic Biototoxin-associated Illness Paradigm to*
680 *Indoor Toxicogenic Fungi, Bioaerosols, Fungi, Bacteria, Mycotoxins and Human*
681 *Health*, Fungal Research Group Foundation, Albany, NY, 2005, pp. 66–77.
- 682 [47] R.C. Shoemaker, P.C. Giclas, C. Crowder, D. House, M.M. Glovsky, Complement split
683 products C3a and C4a are early markers of acute Lyme disease in tick bite patients
684 in the United States, *Int. Arch. Allergy Immunol.* 146 (2008) 255–261.
- 685 [48] R.C. Shoemaker, D.E. House, Sick building syndrome (SBS) and exposure to water-
686 damaged buildings: time series study, clinical trial and mechanisms, *Neurotoxicol.*
687 *Teratol.* 28 (2006) 573–588.
- 688 [49] A.C. Steere, W. Klitz, E.E. Drouin, B.A. Falk, W.W. Kwok, G.T. Nepom, L.A. Baxter-
689 Lowe, Antibiotic-refractory Lyme arthritis is associated with HLA-DR molecules
690 that bind a *Borrelia burgdorferi* peptide, *J. Exp. Med.* 203 (2006) 961–971.
- 691 [50] W. Sun, J. Hong, Y.C. Zang, X. Liu, J.Z. Zhang, Altered expression of vasoactive
692 intestinal peptide receptors in T lymphocytes and aberrant Th1 immunity in
693 multiple sclerosis, *Int. Immunol.* 18 (2006) 1691–1700.
- 694 [51] Y.-R. Tan, T. Yang, S.-P. Liu, Y. Xiang, F. Qu, H.-J. Liu, X.-Q. Qin, Pulmonary
695 peptidergic innervation remodeling and development of airway hyperrespon-
696 siveness induced by RSV persistent infection, *Peptides* 29 (2008) 47–56.
- 697 [52] S.K. Tschoeke, W. Ertel, Immunoparalysis after multiple trauma, *Injury* 38 (2007)
698 1346–1357.
- 699 [53] P. Van Lint, C. Libert, Chemokine and cytokine processing by matrix metallopro-
700 teinases and its effect on leukocyte migration and inflammation, *J. Leukoc. Biol.* 82
701 (2007) 1375–1381.
- 702 [54] R. Wallis, A.W. Dodds, D.A. Mitchell, R.B. Sim, K.B. Reid, W.J. Schwaeble, Molecular
703 interactions between MASP-2, C4, and C2 and their activation fragments leading
704 to complement activation via the lectin pathway, *J. Biol. Chem.* 282 (2007)
705 7844–7851.
- 706 [55] M. Yadav, J. Rosenbaum, E.J. Goetzl, Cutting edge: vasoactive intestinal peptide
707 (VIP) induces differentiation of Th17 cells with a distinctive cytokine profile,
708 *J. Immunol.* 180 (2008) 2772–2776.